Note

Complete Assignment of the ¹H and ¹³C NMR Spectra of Thioquinanthrene and Isothioquinanthrene[†]

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Received 6 January 1997; revised 10 July 1997; accepted 9 August 1997

ABSTRACT: The proton and carbon chemical shifts and the coupling constants "J(H,H) and "J(C,H) of thioquinanthrene and isothioquinanthrene were completely assigned from COSY, HETCOR and INEPT studies. © 1998 John Wiley & Sons, Ltd.

KEYWORDS: NMR; ¹H NMR; ¹³C NMR; 1,4-dithiinodiquinolines; 3- and 4-quinolinyl sulfides

INTRODUCTION

Although thioquinanthrene (1) has been known for about 100 years, the determination of its structure caused problems from the beginning. Additional objections arose when its isomer, isothioquinanthrene (2) was isolated from the reaction mixture. Only in the 1980s was 1 assigned as 1,4-dithiino [2,3-c;5,6-c'] diquinoline and 2 as 1,4-dithiino [2,3-c;6,5-c'] diquinoline.

The simple ¹H NMR spectra of 60 MHz of both dithins 1 and 2 as quinoline derivatives showed two types of signals: a singlet signal of the α -quinolinyl proton (the H-6/H-13 in 1 and the H-6/H-8 in 2), and a multiplet assigned to the remaining protons.⁴ Both

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spectra proved dithiins 1 and 2 to be 3,4-disubstituted quinolines. Nevertheless, some doubts still remained.⁵ The structure of dithiin 1 was definitely confirmed by x-ray determination (using thioquinanthrene dihydrochloride)⁶ and that of isomer 2 using the 1,4-dithiin ring-opening reactions with nucleophiles.⁷

The present study deals with the complete ¹H and ¹³C NMR assignments and the distinction of the two dithiins on the basis of ¹H and ¹³C NMR spectra.

RESULTS AND DISCUSSION

The simple ¹H NMR spectra at 500 MHz of dithiins 1 and 2 showed one singlet of H-6 and four well separated multiplets of four benzene ring protons [H-1, H-2, H-3 and H-4 (both ¹H and ¹³C NMR spectra indicated the symmetry of both dithiins because the number of proton and carbon signals was half the number of these nuclei; to simplify the discussion, only the right part of both molecules, possessing the same numeration, was used in the discussion; Table 1 contains the complete assignment)]. The ¹³C NMR spectra (125 MHz) of 1 and 2 showed five tertiary carbon signals (C-1, C-2, C-3, C-4 and C-6) and four quaternary carbon signals (C-4a, C-6a, C-14a and C-14b).

In order to assign unquestionably all of these signals we used 2D NMR techniques: COSY, HETCOR and INEPT. The HETCOR spectrum permitted the assignment of the tertiary carbon signals and the proton signals via the one-bond proton-carbon correlation. The INEPT spectrum made possible the assignment of the quaternary carbon signals via the three-bond proton-carbon correlation.

The COSY spectrum made it possible to segregate the benzene proton signals as two multiplets with two *ortho* couplings (H-2 and H-3) and two other multiplets with one *ortho* coupling (H-1 and H-4). The distinction between H-1 and H-4 was previously assigned at 80

[†] Part XLV in the series Azinyl Sulfides.

Dithiin	Proton	Carbon single-bond coupling	Carbon two-bond coupling	Carbon three-bond coupling
1	H-1/H-8 (8.37)	C-1/C-8 (123.7)		C-3/C-10 (130.3) C-4a/C-11a (147.2) C-7a/C-14a (144.2)
	H-2/H-9 (7.69)	C-2/C-9 (128.0)		C-4/C-11 (130.0) C-7b/C-14b (126.8)
	H-3/H-10 (7.77)	C-3/C-10 (130.3)		C-1/C-8 (123.7) C-4a/C-11a (147.2)
	H-4/H-11 (8.13)	C-4/C-11 (130.0)		C-2/C-9 (128.0) C-7b/C-14b (126.8)
	H-6/H-13 (8.89)	C-6/C-13 (148.0)	C-6a/C-13a (126.7)	C-4a/C-11a (147.2) C-7a/C-14a (144.2)
2	H-1/H-13 (8.44)	C-1/C-13 (123.5)		C-3/C-11 (130.0) C-4a/C-9a (147.1) C-13b/C-14a (141.6
	H-2/H-12 (7.66)	C-2/C-12 (127.9)		C-4/C-10 (130.1) C-13a/C-14b (126.9
	H-3/H-11 (7.74)	C-3/C-11 (130.0)		C-1/C-13 (123.5) C-4a/C-9a (147.1)
	H-4/H-10 (8.10)	C-4/C-10 (130.1)		C-2/C-12 (127.9) C-13a/C-14b (126.9)
	H-6/H-8 (8.86)	C-6/C-8 (148.1)	C-6a/C-7a (128.9)	C-4a/C-9a (147.1)

Table 1. Complete proton and carbon chemical shifts and summary of HETCOR single-bond and INEPT long-range correlations of dithiins 1 and 2

MHz with the help of 2,4,9,11-tetradeutero-thioquinanthrene (1a).8 It could also be more conveniently performed by means of a three-bond proton—carbon ${}^3J(C,H)$ correlation, as the H-1 proton correlates with three carbons, C-3, C-4a and C-14a, but the H-4 proton correlates with only two carbons, C-2 and C-14b. The identification of the H-1 proton permits complete assignment of benzene ring protons. The long-range correlations H-1/C-14a and H-1, H-3 and H-6/bridged quaternary C-4a and also H-2 and H-4/C-14b confirm a connecting link between the pyridine and benzene parts of 1 and 2, as outlined in Table 1.

The H-6 proton shows ${}^3J(\text{C,H})$ correlations with quaternary carbon resonances corresponding to C-4a and C-14a (at 147.2 and 144.2 ppm, respectively) and ${}^2J(\text{C,H})$ correlation corresponding to C-6a (126.7 ppm). Even though the two-bond C-H couplings ${}^2J(\text{C,H})$ are usually small, the two-bond H-6/C-6a coupling in 1 and 2 (8.3 Hz) is comparable to ${}^3J(\text{C,H})$. Similar effects have been noted with some polycyclic arenes, ${}^{9-11}$ sixmembered aza-hetarenes ${}^{9-11}$ and thienopyridines. 10

The assignment of proton and carbon atoms in dithiin 2 was made analogously to dithiin 1.

One would expect to observe changes in the positions of protons and carbons in the NMR spectra of dithiins 1 and 2 being in the environment of the 3- and 4-sulfide substituents compared with quinoline. However, the H-6 proton signal in both dithiins was almost unaffected ($\delta_{\rm quinoline}=8.92~{\rm ppm^{12}}$), but the C-6 carbon signals were shifted upfield by $\Delta\delta=2.6-2.7~{\rm ppm}$ in comparison with the signal in quinoline (150.7 ppm¹³) Downfield shifts of the signals of C-6a and C-14a (i.e. 3-and 4-quinolinyl) for 1 (126.7 and 144.2 ppm,

respectively) and for 2 (128.9 and 141.6 ppm, respectively) were observed. The differences in magnitude of the deshielding effects are attributable, in our opinion to the differences in the boat conformations of the 1,4-dithiin ring.

C-13b/C-14a (141.6)

The most influenced protons are 5-quinolinyl, i.e. H-1, being shifted downfield in thioquinanthrene (1) by $\Delta \delta = 0.59$ ppm and in isothioquinanthrene (2) by $\Delta \delta = 0.66$ ppm (in comparison with quinoline, 7.78 ppm¹²). In our opinion, the greater deshielding effect in 2 than in 1 is caused not only by the steric interaction between the S-14···H-1 and S-14···H-13 atoms (the peri effect) but also by additional steric interaction between the H-1···H-13 atoms. The same interactions have been observed for other heterocyclodiquinolines.¹¹ The different deshielding effects appear fundamental for the distinction of the two dithins. It is worth nothing that other known pairs of isomeric pentacyclic 1,4dithiinodiazines with a linear fused ring system, 1,4dithiinodiquinolines (2,3-disubstituted quinolines)¹⁴ and 1,4-dithiinodipteridines,¹⁵ showed identical ¹H NMR spectra. Whereas the peri-influenced H-1 atoms were deshielded, the carbon atoms connected with them were shielded ($\Delta \delta = 4.7$ ppm for 1 and $\Delta \delta = 4.9$ ppm for 2) in comparison with the appropriate signals in quinoline.

The chemical shifts of the C-1, C-6, C-6a, C-14a and C-14b carbons in both dithins calculated from the incremental effects of two quinolinylthio substituents, taken separately from the model compounds (3,3'-, 3,4'- and 4,4'-diquinolinyl sulfides), do not confirm the additivity law and suggest steric constraints in the 1,4-dithiin ring.

Whereas all the coupling constants $^{n}J(H,H)$ were

 $^{3}J(H,H)$ $^{1}J(C,H)$ $^{3}J(C,H)$ H,H 2 C,H 2 C,H 2 1 1 1 7.7 7.6 1,2 8.3 8.3 1,1 162.0 161.8 1,3 2,3 6.9 6.9 2,2 162.3 162.1 2,4 8.5 8.5 8.9 3,4 8.4 3,3 161.9 8.4 161.7 3,1 8.8 $^{4}J(H,H)$ 4,4 161.1 163.6 4,2 7.1 7.2 2 H,H 184.4 184.6 6.0 6.2 6,6 4a,1 1.4 1,3 $^{2}J(C,H)$ 4a,3 9.7 9.3 1.4 1.3 C,H 2 12.1 2,4 1.3 4a,6 12.4 $^{5}J(H,H)$ 8.3 8.3 4.9 5.0 6a,6 14a.1 H,H 2 14a,6 6.3 6.2 1 0.7 0.7 1,4 14b,2 8.8 8.7 14b,4 5.2 5.3

Table 2. Coupling constants $^{n}J(H,H)$ and $^{n}J(C,H)$ (Hz) for dithiins 1 and 2

identical for both dithiins, ${}^{1}J(C,H)$ and ${}^{3}J(C,H)$ were slightly different (Table 2).

EXPERIMENTAL

Materials

Compounds 1^{16} and 2^{7} were prepared as described previously.

Spectra

Proton and carbon NMR spectra of 1 and 2 were measured at 27 °C on a Bruker AM 500 spectrometer operating at a proton frequency of 500.13 MHz and a carbon frequency of 125.76 MHz. Approximately 5 mg (for 1 H and 13 C NMR) or 15 mg (for two-dimensional experiments) of each sample were dissolved in 0.5 ml of deuterochloroform. The chemical shifts were referenced to tetramethylsilane. The proton spectra were obtained using an 8 μ s (60°) pulse and a 4 s acquisition time to ensure accurate integrals; 96 transients with 65 536 data points each were used. The 13 C NMR spectra were obtained using a 3 μ s (45°) pulse and a 1.1 s acquisition time to ensure accurate integrals; 272 transients with 65 536 data points each were used.

The homonuclear ${}^{1}H^{-1}H$ two-dimensional correlated diagrams were obtained using the COSY-90 pulse sequence (using a program in the Bruker software). The spectral widths were 440 Hz. The spectra were collected as 25×32 blocks of data, three times zero-filled in the F_1 dimension to obtain the final matrix of 256×128 words. Other parameters were as follows: number of increments in t_1 , 32; number of scans, 8; phase cycling, 8; and relaxation delay, 1.5 s.

The one-bond heteronuclear correlation (HETCOR) spectra were obtained using the XHCORRD program in the Bruker software. The spectra were collected using an acquisition time of 0.27 s, a pulse angle of 90°, a digital resolution of 3.7 Hz, an average ¹J(C,H) of 150

Hz, 32 increments with 8 transients and data processed as a 2048 \times 128 matrix. In the F_1 dimension, the sweep was 350 Hz and the data point resolution was 5.5 Hz per point.

The long-range heteronuclear CH correlation spectra were obtained using the standard pulse sequence (SPT INEPT in the Bruker software). The spectral widths were 30 kHz, the acquisition time was 1.1 s and the relaxation delay was 1.5 s; 320 transients were used per pulsed proton multiplet. Delays for methine protons were adjusted for 8 Hz, giving $D_2 = 11$ ms and $D_3 = 21$ ms, with processing as for the ¹³C NMR spectrum. The value of 8 Hz (or 10 Hz for H-6/C-4a correlation) was used for the coupling.

Selective heterodecoupling (SFDEC in the Bruker software) was used for the $^{n}J(C, H)$ determination.

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